



Molecular Oncology Testing Requisition Form *For internal use in Surgical Pathology ONLY.*

Internal Use Only – Accession #: _____

To Order Oncology Tests:

For Paraffin Embedded Tissue samples, fax completed form to Surgical Pathology at (212)305-2301
For Leukemia and other samples, fax completed form to the PGM Laboratory at (212)342-0420

A. PATIENT INFORMATION:			
LAST NAME:	FIRST NAME:	M.I.:	MRN:
SURGICAL PATHOLOGY REPORT ATTACHED: <input type="checkbox"/> YES (if yes, skip to box D) <input type="checkbox"/> NO			

B. ORDERING PHYSICIAN INFORMATION (OTHER THAN PATHOLOGIST):		
PHYSICIAN LAST NAME:	PHYSICIAN FIRST NAME:	NPI #:
INSTITUTION:	DEPARTMENT:	
ADDRESS:	CITY, STATE & ZIP:	
TELEPHONE NUMBER:	FAX NUMBER:	EMAIL ADDRESS:

C. TISSUE / SAMPLE INFORMATION:	
<input type="checkbox"/> TUMOR - PATHOLOGY SPECIMEN ID NUMBER: _____	Tumor Specimen Type: <input type="checkbox"/> FFPE <input type="checkbox"/> Frozen Tissue
If FFPE, # of slides: _____	Frozen Slides Available in Tissue Bank: <input type="checkbox"/> YES <input type="checkbox"/> NO
Normal (when applicable): <input type="checkbox"/> YES <input type="checkbox"/> NO	Normal Specimen Type /Source: _____
DATE SPECIMEN COLLECTED:	TIME: _____ AM _____ PM
DATE ORDERED: _____	
Ordering Pathologist: _____	Pathologist's Signature _____
Pathologist's Name (Printed)	

D. CLINICAL INFORMATION:	
DIAGNOSIS:	NEOPLASTIC CONTENT:

E. TEST ORDERED (FILL IN COMPLETELY):	
SINGLE GENE TESTING	HEMATOLOGY/ONCOLOGY TESTING
<input type="checkbox"/> BRAF V600E Test <input type="checkbox"/> EGFR Exon 18-21 <input type="checkbox"/> EGFR Expression Test for EGFRvIII <input type="checkbox"/> KRAS Exon 2 <input type="checkbox"/> MGMT Methylation Assay <input type="checkbox"/> MSI Microsatellite Instability Testing	<input type="checkbox"/> Immunoglobulin Heavy Chain (IGH) Gene Rearrangement by Fluorescent PCR <input type="checkbox"/> TCR-beta Rearrangement by Fluorescent PCR <input type="checkbox"/> JAK2 V617F Mutation Test by RT-PCR <input type="checkbox"/> BCR-ABL Quantitative Test by RT-PCR <input type="checkbox"/> IDH1/IDH2 <input type="checkbox"/> Storage Molecular Oncology – HemePath ONLY-no testing performed
NEXT-GENERATION SEQUENCING	
<input type="checkbox"/> TruSeq Amplicon Cancer Panel <input type="checkbox"/> Lung Panel – EGFR, KRAS, BRAF, MET, ERBB2, PIK3CA <input type="checkbox"/> Colorectal Panel – KRAS, BRAF, NRAS, PIK3CA <input type="checkbox"/> GIST Panel – KIT, PDGFRA <input type="checkbox"/> Melanoma Panel – NRAS, BRAF, GNA11, GNAq, KIT <input type="checkbox"/> Cholangiocarcinoma Panel – KRAS, BRAF, NRAS, IDH1 <input type="checkbox"/> Thyroid Panel – AKT1, BRAF, CTNNB1, GNAS, HRAS, KRAS, NRAS, PIK3CA, PTEN, RET, TP53 Complete list of genes on page 2	<input type="checkbox"/> Extended RAS Panel for Thyroid by NGS <input type="checkbox"/> Cancer Whole Exome Sequencing with Transcriptome* <input type="checkbox"/> Columbia Combined Cancer Panel (CCCP)
*Transcriptome is only available on frozen tissue.	



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COLUMBIA COMBINED CANCER PANEL (CCCP) EXONS ONLY

AKT1	BMPRI1A	CDC45	EPHA3	FLCN	IDH1	KMT2C	MRE11A	PDGFRA	PTPRD	SDHB	STAT3	WRN
AKT2	BRCA1	CEBPA	EPHA5	FLT1	IDH2	KMT2D	MSH2	PDPK1	PTPRS	SDHC	STAT5B	WT1
AKT3	BRCA2	CHEK1	EPHB1	FLT3	IFNGR1	KRAS	MSH6	PHF6	PTPRT	SDHD	STAT6	XIAP
ALOX12B	BRCC3	CHEK2	ERBB2	FLT4	IGF1	LAMB4	MTOR	PHF8	PTTG1	SETBP1	STK11	XPA
AMER1	BRIP1	CNOT3	ERBB3	FOXA1	IGF1R	LATS1	MUTYH	PHOX2B	RAC1	SETD2	STK40	XPC
APC	BTX	CREBBP	ERBB4	FOXL2	IGF2	LATS2	MYC	PIGA	RAD21	SF1	SUFU	XPO1
AR	BUB1B	CRKL	ERCC2	FUBP1	IKBKE	LMO1	MYCL	PIK3C2G	RAD50	SF3A1	TP53BP1	YAP1
ARAF	CALR	CRLF2	ERCC3	FYN	IL10	LUC7L2	MYCN	PIK3C3	RAD51	SF3B1	TBL1XR1	YES1
ARID1A	CARD11	CSF1R	ERCC4	GATA1	IL6ST	MAP2K1	MYD88	PIK3CA	RAD51B	SH2B3	TBX3	ZRSR2
ARID1B	CASP8	CSF3R	ERCC5	GATA2	IL7R	MAP2K2	MYOD1	PIK3CB	RAD51C	SH2D1A	TCF3	
ARID2	CBLB	CTCF	ESR1	GATA3	INPP4A	MAP2K4	NBN	PIK3CD	RAD51D	SHQ1	TERT	
ARID5B	CBLC	CTLA4	EXT1	GMNN	INPP4B	MAP3K1	NCOR1	PIK3CG	RAD52	SMAD2	TET1	
ASXL1	CCND3	CUL3	EXT2	GNA11	INSR	MAP3K13	NF1	PIK3R1	RAD54L	SMAD3	TET2	
ASXL2	CCNE1	CYLD	EZH2	GNA13	IRF1	MAPK1	NF2	PIK3R2	RASA1	SMAD4	TET3	
ATM	CD276	DAXX	FAM175A	GNAQ	IRF4	MAX	NFE2L2	PIK3R3	RB1	SMARCA4	TGFBR1	
ATR	CD58	DCUN1D1	FAM46C	GNAS	IRF8	MCL1	NIPBL	PLK2	RBM10	SMARCB1	TGFBR2	
ATRX	CD6	DDB2	FANCA	GNB1	IRS1	MCM2	NKX2-1	PMAIP1	RECQL4	SMARCD1	TMEM127	
AURKA	CD7	DDR2	FANCC	GOPC	IRS2	MCM3	NKX3-1	PMS1	REL	SMARCE1	TNFAIP3	
AURKB	CD73	DICER1	FANCD2	GREM1	JAK1	MCM4	NOTCH1	PMS2	RFWD2	SMC1A	TNFRSF14	
AXIN1	CDC45	DIS3	FANCE	GRID1	JAK3	MCM5	NOTCH2	PNRC1	RHOA	SMC3	TOPBP1	
AXIN2	CDH1	DNM2	FANCF	GRIN2A	JUN	MCM6	NOTCH3	POLE	RICTOR	SMO	TP53	
AXL	CDK12	DNMT1	FANCG	GSK3B	KCNJ5	MCM7	NOTCH4	POT1	RIT1	SOCS1	TP63	
B2M	CDK4	DNMT3A	FAS	H3F3A	KDM5C	MDC1	NRAS	PPP2R1A	RNF43	SOX17	TRAF7	
BAP1	CDK8	DNMT3B	FAT1	H3F3C	KDM6A	MDM2	NTSC2	PRDM1	RPL10	SOX2	TSC1	
BARD1	CDKN1A	DOT1L	FBXO11	HGF	KDM6B	MDM4	NTRK2	PRF1	RPL5	SOX9	TSC2	
BBX3	CDKN1B	E2F3	FBXW7	HIST1H1C	KDR	MED12	PAK1	PRPF40B	RP56KA4	SPEN	TSHR	
BCL11B	CDKN2A	ECTZL	FGF19	HIST1H2BD	KEAP1	MEF2B	PAK7	PRPF8	RP56KB2	SPOP	U2AF1	
BCL2L1	CDKN2B	EED	FGF3	HIST1H3B	KIAA1549	MEN1	PALB2	PTCH1	RPTOR	SRC	U2AF2	
BCL2L11	CDKN2C	EGFL7	FGF4	HNF1A	KIT	MET	PARK2	PTEN	RYBP	SRSF2	UBR5	
BCL6	CDT1	EGFR	FGFR2	HRRAS	KLF4	MITF	PARP1	PTPN1	SBD5	STAG1	VHL	
BCORL1	CD79A	E1F1AX	FGFR4	ICOSLG	KLF6	MLH1	PBRM1	PTPN11	SDHA	STAG2	VTCN1	
BLM	CD79B	EPCAM	FH	ID3	KLHL6	MPL	PDCD1	PTPRC	SDHAF2	STAG3	WAS	

COLUMBIA COMBINED CANCER PANEL (CCCP) WHOLE GENES

ABL1	BRD4	CIITA	EP300	EWSR1	FUS	KAT6A	NPM1	PAX5	PPARG	RUNX1	SUZ12	TPM3
ALK	CBL	CLTC	ERG	EZR	GPC3	KIF5B	NTRK1	PAX8	PRKAR1A	SLC45A3	SYK	USP6
BCOR	CD274	CLTCL1	ETV1	FGFR1	HMGA2	KMT2A	NTRK3	PDGFRB	RAF1	SS18	TAF15	
BCR	CD74	CREBBP	ETV4	FGFR3	IL2	LRI3	NUP214	PICALM	RARA	SSX1	TCF12	
BRAF	CDK6	CRLF2	ETV5	FIP1L1	ITK	MECOM	NUP98	PLAG1	RET	SSX2	TFE3	
BRD3	CIC	CTNBN1	ETV6	FOXO1	JAK2	MLL2	NUTM1	PML	ROS1	SSX4	TMPRSS2	

TRUSeq AMPLICON CANCER PANEL

ABL1	ATM	CSF1R	ERBB4	FGFR3	GNAS	JAK2	KRAS	NOTCH1	PIK3CA	RET	SRC
AKT1	BRAF	CTNBN1	FBXW7	FLT3	HNF1A	JAK3	MET	NPM1	PTEN	SMAD4	STK11
ALK	CDH1	EGFR	FGFR1	GNA11	HRRAS	KDR	MLH1	NRAS	PTPN11	SMARCB1	TP53
APC	CDKN2A	ERBB2	FGFR2	GNAQ	IDH1	KIT	MPL	PDGFRA	RB1	SMO	VHL

INSTRUCTIONS FOR SUBMISSION OF MOLECULAR ONCOLOGY SPECIMENS

BRAF: 10 unstained, unbaked sections on regular slides with a serial H&E-stained section to document presence of tumor in block. If the section clearly contains greater than 40% tumor cells (not tumor area), then five 10-micrometer sections in a 1.5-2ml DNase-free, RNase-free microcentrifuge tube along with a serial H&E-stained section will be accepted.

Cancer Whole Exome Sequencing with Transcriptome (CWES): Performed only on snap frozen tissue in the CLIA tissue bank. 10 unstained slides and 1 H&E are required from the tumor. A normal control from the patient is also required (blood or buccal swabs) and should be submitted to PGM by the clinical team.

EGFR and/or KRAS Mutation Test by Sequencing: 10, 10-micrometer blanks from paraffin block containing highest percentage of tumor, on non-coated slides; do not bake slides. Submit one H&E stained-slide from same paraffin block, cut after the 5th blank (note: if ordering EGFR and KRAS mutation on same paraffin block, only one sample is needed).

EGFRviii Tumor Sample: 5-10 10-µm sections of tumor in a 1.7ml or 2.0 ml microcentrifuge tube. Label tube with surgical pathology accession number and block designation. **One H&E-stained slide:** Cut either immediately prior to or after paraffin sections above ("serial section"). In lieu of a serial section, a notation from a neuropathologist, indicating that the sections contain greater than 50% tumor, is also acceptable. Submit to lab at room temperature.

MGMT Tumor Sample: 5-10 5-µm sections of tumor in a 1.7ml or 2.0 ml microcentrifuge tube. Label tube with surgical pathology accession number and block designation. **One H&E-stained slide:** Cut either immediately prior to or after paraffin sections above ("serial section"). (In lieu of a serial section, a notation from a neuropathologist, indicating that the sections contain greater than 50% tumor, is also acceptable. Submit to lab at room temperature. (1-5 unstained slides in lieu of or tissue in microcentrifuge tube, will also be accepted; if section has less than 50% tumor, please submit slides, and circle area of tumor; do not bake slides). If the original pathology had been performed at CUMC, please fax requisition above to the attention of attending who signed out the original surgical case to have case cut.

MSI Microsatellite Instability: Five 10-µm sections of tumor and normal in separate PCR tubes with serial H&E of tumor. **If less than 50% tumor in block,** then five 10-µm unstained sections on UNCOATED slides with tumor indicated on serial H&E, along with five sections from normal block in PCR tube, if available. Microdissected DNA from tumor and normal if certified by a Pathologist may also be used.

TruSeq Targeted Cancer Panels & Columbia Combined Cancer Panel (CCCP): 10 unstained, unbaked sections on regular slides with a serial H&E-stained section to document presence of tumor in block.