FINE NEEDLE ASPIRATION BIOPSY KIT INSTRUCTIONS

FNA KIT CONTENTS

1. 2 white containers with 95% ethanol for immersion fixation and transportation of direct smears.
2. 2 plastic slide holders for transportation of air-dried direct smears.
3. 1 tube with CytoLyt fluid for needle rinses and/or cell block preparations.
4. 1 container with 10% formalin for cell block preparations and/or ancillary studies.
5. 1 microcentrifuge tube with RNAProtect® Cell Reagent, for molecular test samples (2 ml).

IN ORDER TO INSURE OPTIMAL FNA SAMPLE PREPARATION, PLEASE FOLLOW THESE INSTRUCTIONS:

• GENERAL GUIDELINES:
  o Fill the entire requisition form with special attention to site, side, and size of targeted lesion.
  o Provide pertinent clinical and imaging information.
  o Preferred are alcohol fixed, thin monolayer direct smears on clean glass slides.
  o Label each slide, containers, and microcentrifuge tube with patient’s name and date of birth.

• CELL PRESERVATION AND FIXATION ON DIRECT SMEARS:
  o Immediately after smearing cellular material on a glass slide, wet-fix direct smears by submerging them in 95% ethanol, contained in white containers.
  o Do not allow material to air dry!
  o Clotted material expelled onto slide for direct smears should be placed into formalin (not smeared).
  o Slides must be left in ethanol-filled containers for transportation.
  o Ensure lids are screwed on tight to avoid leakage.

• ADDITIONAL CELLULAR MATERIAL ON DIRECT SMEARS FOR MODIFIED GIEMSA STAIN:
  o Leave 1 or 2 direct smears air-dried and unfixed.
  o Place in plastic slide holders for transportation.

• ANCILLARY STUDIES AND CELL BLOCK PREPARATION:
  o Thoroughly rinse the needle from each pass in tube with CytoLyt.
  o Dedicated needle passes placed directly into formalin are suggested for cellular cell blocks. This enriched cellular material will be utilized for appropriate ancillary studies.
  o Preferred are formalin-fixed and cellular cell blocks in cases where study of tumor markers with prognostic and therapeutic significance is a consideration (i.e. breast, lung, etc.).
  o If flow cytometric evaluation is required to rule out a hematolymphoid neoplasm, submission of aspirate in RPMI is recommended. Call the Lab to request RPMI tubes with 24-48 hrs notice.

• MOLECULAR TESTING:
  o 2 dedicated needle passes should be placed in the microcentrifuge tube.
  o Label tube with patient’s name and DOB.
  o Up to 0.3 ml of material can be added to this tube. The tube has 1.5 ml of fluid, with 0.5 ml up to the next mark on the tube.
  o Immediately turn over the tube 15-20 times to mix the cells with the reagent.

If clarification is needed regarding the above instructions, call Cytology at 212-305-2360.
For additional FNA collection kits, please contact us at:
COLUMBIA UNIVERSITY PATHOLOGISTS
800-653-8200 / 212-305-4840